

Dehydrated Culture Media Bases / Media Supplements

# **Technical Information**

## L. D. Agar

## Product Code: DM 1742

Application: - L. D. Agar is used for cultivation and identification of fastidious anaerobic bacteria.

Composition**				
Ingredients	Gms / Litre			
Casein enzymic hydrolysate	5.000			
Yeast extract	5.000			
Sodium chloride	2.500			
Sodium sulphite	0.100			
L-Cystine	0.400			
L-Tryptophan	0.200			
Vitamin K1	0.010			
Hemin	0.010			
Agar	20.000			
Final pH ( at 25°C)	7.4±0.2			
**Formula adjusted, standardized to suit p	erformance			
parameters				

parameters

### Principle & Interpretation

Anaerobes are the organisms that grow in the absence of oxygen. Depending upon their ability to tolerate oxygen, they are classified as either facultative or obligate anaerobes. The anaerobic gram-negative bacteria are part of the normal flora of the upper respiratory tract, mouth, intestinal tract and urinogenital tract of human and animals. The bile-resistant *Bacteroides fragilis* more resistant to antimicrobial agents is the most commonly recovered anaerobe than any other anaerobe in clinical specimens. *Fusobacterium necrophorum* is a very virulent anaerobe that may cause severe infections, usually in children or young adults <sup>(5)</sup>.

L. D. Medium or Lombard-Dowell Medium was developed by Dowell and Lombard <sup>(1)</sup> for the cultivation and identification of fastidious anaerobic bacteria. L. D. Agar is used to evaluate the degree of growth of anaerobes including indole and catalase production by *Bacteroides* and *Fusobacterium* species isolated from clinical specimens.

L. D. Agar is essentially a casein digest agar enriched with hemin, vitamin K1, L-cystine and yeast extract<sup>(3)</sup>. This medium contains various nutritious substances, which can promote the growth of fastidious anaerobic bacteria. Casein enzymic hydrolysate and yeast extract provide the necessary nitrogenous nutrients while hemin and vitamin K1 supply additional growth factors. L-cystine and L-tryptophan serve as the amino acid sources. Sodium sulphite is an antioxidant. Sodium chloride maintains osmotic balance of the medium. Catala se-positive reaction may not be evident uptill 30 seconds to 1 minute after application of 3% hydrogen peroxide<sup>(2, 4)</sup>.

## Methodology

Suspend 33.22 grams of powder media in 1000 ml distilled water. Shake well and heat to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (12 1°C) for 15 minutes. Mix well and pour into sterile Petri plates.

## **Quality Control**

#### **Physical Appearance**

Cream to yellow homogeneous free flowing powder

Gelling

Firm, comparable with 2.0% agar gel.





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#### Colour and Clarity of prepared medium

Medium amber coloured clear to slightly opalescent gel forms in Petri plates

#### Reaction

Reaction of 3.32% w/v aqueous solution at 25°C. pH : 7.4±0.2

pH Range 7.20-7.60

#### Cultural Response/Characteristics

DM 1742: Cultural characteristics observed under anaerobic condition, after an incubation at 35-37°C for 40-48 hours.

Organism	Growth	Indole production	Catalase
Bacteroides fragilis ATCC 25285	good-luxuriant	negative reaction	positive reaction
Bacteroides corrodens	fair-good	negative reaction	negative reaction
Fusobacterium necrophorum ATCC 25286	good-luxuriant	positive reaction	negative reaction
Fusobacterium nucleatum ATCC 25586	fair-good	positive reaction	negative reaction

### Storage and Shelf Life

**Dried media:** Store below 30°C in tightly closed container and use before expiry date as mentioned on the label. **Prepared Media:** 2-8<sup>0</sup> in sealable plastic bags for 2-5 days.

### **Further Reading**

1. Dowell V. and Lombard G., June 1977, U.S., DHEW, Center for Disease Control (CDC), Atlanta. Ga.

2. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore

3. Finegold S. M., Baron E. J., Bailey and Scotts Diagnostic Microbiology, 8th Ed., 1990, The C.V. Mosby Company

4. Koneman E., Allen S., Dowell V. and Sommers H., 1979, Colour Atlas and Textbook of Diagnostic Microbiology, J. B. Lippincott Co., Philadelphia.

5. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Yolken R. H., (Ed.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology, Washington, D.C.

### **Disclaimer**:

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